Lokiarchaeota marks the transition between the archaeal and eukaryotic selenocysteine encoding systems

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Abstract

Selenocysteine (Sec) is the 21st amino acid in the genetic code, inserted in response to UGA codons with the help of RNA structures, the SECIS elements. The three domains of life feature distinct strategies for Sec insertion in proteins and its utilization. While bacteria and archaea possess similar sets of selenoproteins, Sec biosynthesis is more similar among archaea and eukaryotes. However, SECIS elements are completely different in the three domains of life. Here, we analyze the archaean *Lokiarchaeota* that resolves the relationships among Sec insertion systems. This organism has selenoproteins representing five protein families, three of which have multiple Sec residues. Remarkably, these archaeal selenoprotein genes possess conserved RNA structures that strongly resemble the eukaryotic SECIS element, including key eukaryotic protein-binding sites. These structures also share similarity with the SECIS element in archaeal selenoprotein *Vhud*, suggesting a relation of direct descent. These results identify *Lokiarchaeota* as an intermediate form between the archaeal and eukaryotic Sec-encoding systems and clarify the evolution of the Sec insertion system.
Introduction

2.1 Selenoproteins across the domains of life

Selenoproteins are a rare class of proteins that contain a selenocysteine (Sec) residue, often referred to as the 21st amino acid (Labunskyy et al., 2014). Sec is inserted co-translationally, like standard amino acids, and possesses its own tRNA (tRNAsec). However, Sec does not have a fully dedicated codon in the genetic code. Instead, Sec residues are inserted in response to UGA codons that are recoded in the presence of specific Sec designation signals; UGAs are otherwise interpreted as stop signals in most organisms. In response to those signals, a Sec specific elongation factor (EFsec) replaces the standard EF-Tu uniquely for the translation of Sec UGA codons. EFsec recruits the Sec-tRNA, promoting the specific insertion of Sec residues at these locations.

Most selenoproteins are enzymes with oxidoreductase function, with Sec being the catalytic redox active site. For the great majority of selenoproteins, standard homologues (orthologues and/or paralogues) that replace Sec with cysteine (Cys) exist, and are known to perform essentially the same molecular function, albeit less efficiently (Fomenko and Gladyshev, 2012). Although the exchangeability of Sec and Cys is debated (Castellano, 2009; Gromer et al., 2003; Hondal and Ruggles, 2011; Hondal et al., 2013), it is generally accepted that Sec is used instead of Cys for reasons related to its higher reactivity, which leads to improved catalytic efficiency or resistance to inactivation in redox reactions. Many selenoproteins are enzymes involved in redox homeostasis, including some that are essential for human, mouse and other vertebrates (Labunskyy et al., 2014).
Selenoproteins require a specific set of genes dedicated to Sec synthesis and insertion (here denoted as the "Sec machinery"), including EFsec and tRNAsec. However, selenoproteins, along with the capacity to code for Sec (the “Sec trait”), are not present in all organisms. Their distribution is scattered, but encompasses lineages of the three domains of life: bacteria, archaea, and eukaryotes (Mariotti et al., 2015). Selenoprotein families are quite diverse, with little overlap between prokaryotic and eukaryotic selenoproteomes (Driscoll and Chavatte, 2004). Bacteria utilize selenoproteins to carry out functions such as redox homeostasis, electron transport / energy metabolism, compound detoxification and oxidative protein folding. In contrast, the archaeal selenoproteins with known functions are involved in hydrogenotrophic methanogenesis, with the only exception of selenophosphate synthetase (SPS), involved in Sec biosynthesis (Stock and Rother, 2009). Eukaryotic selenoproteins, on the other hand, carry out very diverse functions. In comparison to prokaryotes, eukaryotic selenoproteomes are largely expanded in families involved in redox regulation, antioxidant defense, protein repair and oxidative protein folding, while they are decreased or depleted in families involved in compound detoxification, electron transport and energy metabolism (Labunskyy et al., 2014).

The mechanisms of Sec synthesis and insertion also exhibit differences in the three domains of life (Figure 1). However, while archaea and bacteria share a larger number of selenoprotein families than with eukaryotes, Sec biosynthesis is more similar between archaea and eukaryotes. Organisms in these two lineages first catalyze the synthesis of phospho-Ser with the protein PSTK, and then convert it to Sec, whereas bacteria directly synthesize Sec from Ser.

2.2 SECIS elements

The main signals for Sec insertion are RNA structures found in the selenoprotein transcripts, designated SECIS (SEC Insertion Sequence) elements (Berry et al., 1991). Strikingly, SECIS elements do not share any obvious resemblance in sequence or structure between the three
domains of life (Krol, 2002). In bacteria, the SECIS element (bSECIS) is a stem-loop structure located within the coding sequence, immediately downstream of the Sec UGA site it acts upon (Figure 1B) (Hüttenhofer et al., 1996). These structures show considerable variability across genes and species, but always feature at least one stem and a small loop, with one or more G in the first loop positions (Zhang and Gladyshev, 2005). The recognition of bSECIS elements is carried out by a specialized domain of the bacterial EFsec (SelB) (Yoshizawa and Böck, 2009).

The eukaryotic SECIS elements are located in the 3’ UTR of selenoprotein transcripts. The distance between the SECIS and the Sec UGA codon varies substantially in vertebrates (~0.2 / 5.2 kB (Mariotti et al., 2012)). Eukaryotic SECISes contain an RNA motif called kink-turn (Latrèche et al., 2009), characterized by an unusual GA-GA antiparallel pair, and surrounded by two stems. An additional stem is found in the apical loop of certain SECIS elements, known as “type II” (Grundner-Culemann et al., 1999). Eukaryotic SECISes show a few other conserved features besides the invariant kink-turn core. The dinucleotide preceding the first GA is strongly conserved as AU (forming the AUGA tetranucleotide), or rarely substituted with GU. The first nucleotides in the loop at the top of stem 2 are strongly conserved as adenines, with few exceptions. Lastly, additional positions around the core show preference towards specific nucleotides (Chapple et al., 2009). Eukaryotic SECISes are bound by SBP2, a L7AE-domain containing protein that contacts the region around the core (Fletcher et al., 2001). Besides recognizing SECIS elements, SBP2 interacts with EFsec at the Sec UGA site (Tujebajeva et al., 2000).

In archaea, SECIS elements (aSECIS) are also located in the 3’ UTR of selenoprotein genes, with a single documented exception in which it is found in the 5’ UTR (Wilting et al., 1997). Archaeal SECISes (Krol, 2002; Kryukov and Gladyshev, 2004; Stock and Rother, 2009) are characterized by two stems separated by an invariant asymmetric bulge, consisting of a GAA
trinucleotide at the 5' and a single adenine at the 3'. The first stem is GC rich and encompass 10 base pairs. The second stem is shorter, generally only 3 base pairs, and is entirely composed of GC pairs. The apical loop has variable length, and may contain additional pairings. To date, the archaeal SECIS element has no known interactor protein, and the question of how the SECIS and the Sec UGA site communicate remains open. The SBP2 counterpart has never been observed in archaea, while the archaeal EFsec was shown not bind archaeal SECIS elements (Stock and Rother, 2009).

2.3 Evolution of the Sec trait

The distribution of selenoproteins in living organisms, particularly in prokaryotes and protists, testifies a dynamic process in which the Sec trait was lost in many lineages independently (Zhang et al., 2006; Lobanov et al., 2008; Mariotti et al., 2015). Although some isolated events of horizontal gene transfer have occurred, in general both the distribution of proteins in the Sec pathway and their reconstructed phylogeny are consistent with the known phylogenetic relationship between bacteria, archaea and eukaryotes (Mariotti et al., 2015). In addition, considering the evident homology of the core Sec pathway (tRNAsec, EFsec, SPS) it becomes compelling that the Sec trait has evolved only once in the history of life, and its origin can be dated back to the last universal common ancestor. Yet, SECIS elements bear no obvious resemblance between the major domains of life. This is particularly surprising when we consider archaea and eukaryotes, in which SECIS elements are situated in the same location (the 3'UTR), and are thus expected to be homologous structures.

We must consider, however, that selenoproteins are a very rare feature among the archaea analyzed so far. The markers for Sec utilization were identified only in two phylogenetic orders, Methanococcales and Methanopyrales (Methanopyrus), while all other major archaeal groups seem to be devoid of selenoproteins (Stock and Rother, 2009). Since the time of the discovery
of archaeal selenoproteins (Wilting et al., 1997), many new archaeal sequences became available, but, until today, the Sec trait has remained limited to the same two genera.

2.4 The origin of eukaryotes and the discovery of Lokiarchaeota

Eukaryotes possess several features that set them apart from prokaryotes, such as a larger cell size and a complex cell compartmentalization (including the presence of the nucleus, mitochondria and cytoskeleton). Remarkably, these complex features are present in nearly all extant eukaryotes without any apparent intermediate grade, posing the problem of how the first eukaryotes came about. In recent years, the increasing availability of genome sequences and the development of sophisticated phylogenomics tools has finally shed light into the origin of this domain of life. It is now widely accepted that eukaryotes originated from an archaeal ancestor, and that a key event was the incorporation of an alpha-proteobacterial endosymbiont, giving rise to mitochondria (Koonin, 2010). The phylogenetic relationships between extant archaea and the proto-eukaryotic archaean ancestor, however, remained uncertain: it was unclear whether eukaryotes should be considered a sister group of archaea, or rather branched from within archaean.

A recent study (Spang et al., 2015) has reported the discovery of a novel archaean phylum, Lokiarchaeota, in marine sediments collected near a hydrothermal vent known as Loki’s Castle. In their work, the authors describe how they obtained a metagenomic assembly of non-amplified DNA from this site (LCGC14, Genbank accession LAZR01000000.1), hereafter referred to as the Laz sequence set. From this assembly, they derived a contig subset by performing supervised binning driven by a carefully selected set of genes with a peculiar phylogenetic signal. The resulting set of contigs (Lokiarchaeum genome bins, Genbank accession JYIM00000000.1), hereafter designated as Loki, was subject of extensive phylogenetic analysis. The Loki sequences turned out to belong to a new archaean lineage, that appears as a sister
group of eukaryotes in phylogenetic reconstructions. This strongly supports the hypothesis that eukaryotes evolved from within the archaeal domain, with *Lokiarchaeota* being the closest known relative to the archaeal ancestor of eukaryotes (Eme and Doolittle, 2015; Finn et al., 2016; Koonin, 2015). Consistent with this, several eukaryotic signature proteins were found in *Lokiarchaeota*.

In this work, we present the first analysis of selenoproteins and Sec pathway in *Lokiarchaeota*. A complete set of known archaeal genes to encode for Sec was found in *Loki*, together with selenoprotein genes belonging to five distinct families. Remarkably, the selenoprotein genes in *Lokiarchaeota* possess conserved RNA structures that resemble the eukaryotic SECIS elements. These results identify *Lokiarchaeota* as an intermediate form between the typical archaeal and eukaryotic Sec encoding systems, contributing to the understanding of the origin and evolution of the Sec insertion system.

**Results**

We analyzed selenoproteins and Sec biosynthesis genes in the *Loki* sequences and further expanded our searches to the *Loki* superset assembly, *Laz*. Both *Laz* and *Loki* are metagenomes, representing a mixture of sequences from a multitude of species (although *Loki* obviously displays a much smaller diversity, approximable to a single genome assembly). Both *Laz* and *Loki* are affected by the potential confounding factor of “apparent paralogues”: genes in multiple copies in the metagenome, which actually correspond to orthologous genes of closely related strains or species. To avoid this confounding issue, we decided to analyze each gene in its genomic context (Methods, Supplementary Material S1). This analysis reduced redundancy of the sets of genes of interest in *Loki*, which allowed us to analyze the sequences in greater detail.
3.1 A complete Sec machinery in *Loki* sequences

We used a variety of computational approaches to search the *Loki* sequences for genes involved in the Sec pathway (Methods). We identified the full set of known genes required for archaeal Sec utilization, here designated as the Sec machinery: *tRNAsec*, *pstk*, *SecS*, *EFsec*, *SPS* (genomic locations are provided in Supplementary Table S1). All Sec machinery genes were found to be present in single copy in *Loki* and on distinct contigs, with the exception of *EFsec* and *tRNAsec*. *EFsec* was present in the same contig with *tRNAsec*, approximately ~6 Kb and 8 genes upstream, on the opposite strand. This pair of genes was observed twice, on two distinct contigs in *Loki*. However, we could classify these as apparent paralogues, belonging to very closely related *Lokiarchaeota*. In fact, in both contigs the homology extended to the region between *tRNAsec* and *EFsec*, and beyond (Supplementary Material S1.A). The only non-protein coding gene in the Sec machinery, *tRNAsec*, contained a 31 nt intron interrupting the loop in the TΨC arm (Figure 2). Although this position is considered not canonical for tRNA introns, it was previously observed in other archaeal tRNAs (Yoshihisa, 2014). The sequence and structure of the mature tRNA resembles unequivocally the archaeal *tRNAsec*. Similarly, all Sec machinery proteins exhibited an archaeal phylogenetic signal, being more similar to *Methanococcales* or *Methanopyrus* orthologues than to their bacterial or eukaryotic counterparts (Supplementary Material S2).

The *SPS* gene (selenophosphate synthetase) is singular in that it is both part of the Sec machinery and a selenoprotein itself in many organisms. *SPS* is an ancestral selenoprotein, and although it has replaced Sec with Cys in many organisms, most likely its original form contained Sec (Mariotti et al., 2015). In *Loki*, we identified only a fragment of the *SPS* gene. It was located on a short contig, which ended in the middle of the *SPS* coding sequence, resulting in an apparently truncated sequence that lacked the Sec-containing N-terminal region. However, we
could deduce the complete *Loki* SPS protein sequence analyzing *Laz* sequences. We found several SPS genes, some of which were almost identical in sequence to the *Loki* SPS fragment. We thus collected all SPS genes in *Laz*, aligned them with a reference set of SPS proteins from the whole tree of life (Mariotti et al., 2015), and applied a phylogenetic reconstruction procedure (Methods). The resulting tree offered a “phylogenetic fingerprint” of all SPS containing species present in *Laz* (Supplementary Material S2.A). While some of the *Laz* SPS genes clustered within diverse bacterial lineages, a group of 9 SPS genes (including the *Loki* sequence fragment) branched within archaea, forming a cluster that resembled the *Methanococcales* SPS gene, yet exhibited considerable sequence divergence. We further refer to these *Laz* genes as the *Loki*-like SPS. Their coding sequence contained one in-frame UGA codon aligned to the homologous Sec position in other species, supporting the existence of SPS as selenoprotein in *Lokiarchaeota*. We also searched the *Loki* and *Laz* assemblies for the presence of the selenouridine synthase (*ybbB* gene), a marker of utilization of selenouridine in certain tRNAs (Romero et al., 2005). However, none of the possible candidates withstood our manual inspection. Likewise, the search for gene markers of Se utilization as cofactor to certain molybdenum-dependent hydroxylases (*YqeB* and *YqeC*) (Lin et al., 2015) did not return any suitable candidate. Thus, the use of Sec in selenoproteins is the only known Se utilization trait supported by SPS in *Lokiarchaeota*.

### 3.2 Selenoprotein families in *Lokiarchaeota*

We applied multiple approaches to search for selenoprotein genes in *Lokiarchaeota* (Methods). These methods were aimed to identify genes belonging both to known selenoprotein families and to novel families. This search yielded *Loki* selenoprotein genes belonging to 5 protein families: SPS, *VhuU*, *VhuD*, *HdrA*, and *PrxL* (Figure 3). Of these, *VhuU*, *VhuD* and *HdrA* were orthologous to the selenoprotein genes previously identified in other archaea genera, such as...
Methanococcus and Methanopyrus (Stock and Rother, 2009), and they clustered together in phylogenetic reconstructions (Supplementary Material S3).

VhuU and VhuD encoded two subunits of the same enzyme, the F$_{420}$-non-reducing hydrogenase (Vhu), which includes also non-selenoprotein subunits VhuA and VhuG (both found in the Loki genome, see Supplementary Table S1). In Loki and in previously described Sec-containing archaea alike, VhuU is a small oxidase (~50 amino acids) with a single Sec residue, while VhuD is a ~150 amino acid-long hydrogenase with two distinct Sec positions (Wilting et al., 1997).

HdrA represents the largest subunit (~660 amino acids) of heterodisulfide reductase (Hdr) complex, which participates, together with Vhu, in the reduction of Coenzyme B-Coenzyme M heterodisulfide (CoM-S-S-CoB) during the last step of methanogenesis. Hdr includes also subunits HdrB and HdrC. These do not contain Sec, and were also identified in the Loki genome (Supplementary Table S1). In the Sec utilizing archaeal species previously described, HdrA has a single Sec residue in position ~200 (“canonical position”). In contrast, in Loki sequences we identified several multi-Sec HdrA genes, with a total of 5 possible Sec positions, always including the canonical one (Figure 3). All these positions aligned to conserved Cys residues in other archaeal HdrA proteins. Strikingly, all Loki HdrA and VhuD genes were located in tandem syntenic pairs, with VhuD always located downstream of HdrA on the same strand (Figure 3). This syntenic block was also conserved in all Methanococcales archaea as well as many other archaeal lineages. The intergenic space between these two genes in Loki is invariably very short. In one case (HdrA.3 - VhuD.3), the two coding sequences actually overlap by 28 bp, in different frames.

The remaining Sec-containing proteins identified in Loki resemble the peroxiredoxin family (Prx), which was never observed as selenoproteins in archaea. Interestingly, these Loki selenoproteins belong to a subclass that lacks typical Prx features. This diverse family, never
experimentally characterized, was previously referred to as Prx-like (Fomenko and Gladyshev, 2003; Cui et al., 2012). Whereas Prx proteins contain an active site with Thr/Ser-X-X-Cys (where X stands for any residue) (Poole and Nelson, 2016), Prx-like proteins possess a Cys-X-X-Cys motif (“redox box”) typical of thioredoxins, with the first Cys replaced by Sec in some bacteria (Zhang and Gladyshev, 2008) and eukaryotes (Jiang et al., 2012; Mariotti et al., 2013).

We identified two distinct Prx-like genes in Loki, hereafter referred to as PrxL. The PrxL.2 gene contains a single Sec residue corresponding to the first Cys in the redox box, while PrxL.1 contains two, thus replacing both Cys residues with Sec. Searching Laz sequences, we identified additional PrxL homologues, either with one or two Sec residues at the same positions (Supplementary Table S1). Strikingly, none of the PrxL proteins contain any additional Cys residue outside the redox box.

3.3 The Loki SECIS element resembles the eukaryotic SECIS

After identifying the selenoprotein genes in Loki, we searched their sequences for the occurrence of SECIS elements. To our surprise, none of the tools designed to identify archaeal, bacterial or eukaryotic SECIS elements reported any significant hit in Loki selenoprotein genes. Hence, we outlined a de novo computational procedure aimed to identify any RNA motif conserved in these genes (Methods). With this, we detected a common RNA structure located downstream of the coding sequences in every Loki selenoprotein gene with the exception of SPS (explained later). Following our initial discovery of the structures in the selenoprotein genes located on the Loki contigs (Supplementary Material S4.A), we expanded the search to the full set of Loki-like selenoproteins in Laz (see Methods), ending up with a final set of 25 different “lokiSECIS” elements (Supplementary Material S4.B). All these structural motifs were located in close proximity to the end of the selenoprotein coding sequences (Figure 3). In all occurrences of the tandem genes HdrA and VhuD, only a single lokiSECIS was found downstream of the second gene (VhuD), but not anywhere else nearby, suggesting that these pairs of genes share
their Sec insertion signal. All lokiSECIS elements in PrxL, VhuU, HdrA-VhuD folded in a very precise shape, featuring a 9-bp stem and an 11-nt apical loop. In a few VhuD SECIS elements, the region corresponding to the apical loop was predicted with additional pairings, forming an additional short stem (Supplementary Material S4.A). From the alignment of all lokiSECIS elements, their conserved features were clearly discernible (Figure 4C).

From the analysis of lokiSECIS elements, we observed the following: (i) the nucleotides preceding the stem were always AUGA; (ii) the first five nucleotides of the apical loop showed a strong preference for adenines; (iii) downstream of the stem, the first two nucleotides were GA (except that they were AA in VhuU). Remarkably, all these characteristics of the lokiSECIS were shared with the eukaryotic SECIS elements. At glance, the lokiSECIS looks like a eukaryotic SECIS with a shorter stem. The lokiSECIS exhibited a few additional conserved features (Figure 4C), the most prominent being an ultraconserved guanine at the last position of the apical loop, most often preceded by an adenine. We detected a lokiSECIS element with these characteristics downstream of every selenoprotein gene in the Loki contigs, as well as downstream of those selenoprotein genes in Laz which we also attributed to Lokiarchaeota, with one exception (SPS genes).

Applying an alternative discovery approach (Methods), we found that the Loki-like SPS genes also possessed a conserved structure in the same homologous position. This structure (Figure 5, Supplementary Material S4.C) also resembled the eukaryotic SECIS, and had some obvious similarities with the other lokiSECIS elements, but showed important differences as well. All lokiSECIS elements of PrxL, VhuU, HdrA-VhuD genes possessed the same number of nucleotides in the stem and apical loop. In contrast, the main stem was shorter in the SPS lokiSECIS, but the overall structure was longer, featuring in all cases an additional stem in the
apical region. In this regard, the SPS lokiSECIS resembled a eukaryotic type II SECIS, except that it had a shorter stem.

Next, we set out to investigate where the lokiSECIS originated from, searching all publicly available archaeal genomes for similar structures using lokiSECISearch (Methods). This analysis resulted in no significant hits in the great majority of genomes (90%), and only one or two hits in each of the remaining genomes (with the sole exception of Loki, in which the program recovered all lokiSECIS elements previously described). However, we noticed that for Methanococci species M.aeolicus and M.vannielii, the unique lokiSECIS hits were located in orthologous positions. Strikingly, they mapped just downstream of the selenoprotein gene VhuD and corresponded to the archaeal SECIS of these genes. To further investigate this, we predicted the archaeal SECIS elements of all selenoprotein genes in Methanococcales and Methanopyrus. We then extended their sequence boundaries beyond the limits normally considered for archaeal SECIS (i.e., Figure 4A), and aligned them. Although this procedure did not uncover new features conserved across all SECIS elements of archaea, we detected motifs conserved in a gene-specific fashion (Supplementary Material S4.D). The archaeal VhuD SECISes were particularly intriguing: they all possessed a short extra stem. Moreover, an invariant AUGA motif on one side, and a GAC/AAC trinucleotide on the other, was observed in between the extra stem and the main stem (Figure 4B). Thus, the Methanococcales and Methanopyrus VhuD SECIS elements exhibit both the typical features of archaeal SECIS elements and the distinctive motifs of the Loki and eukaryotic SECIS.

3.4 SBP2 is not detected in Lokiarchaeota

At present, no SECIS binding protein is known in the Sec utilizing archaea Methanococcales and Methanopyrus. In contrast, it is well established that eukaryotic SECIS elements are recognized by SBP2, a master regulator in the Sec insertion process (Kossinova et al., 2014),
although recent results questioned the absolute requirement of SBP2 for selenoprotein expression (Seeher and Schweizer, 2014). Although with remarkable variability in size, SBP2 is found in every selenoprotein containing eukaryotic genome, and always includes a L7AE domain (Donovan and Copeland, 2009). This domain is shared with other RNA binding proteins that also recognize kink-turn motifs (Huang and Lilley, 2013). In consideration of the similarity of the lokiSECIS with the eukaryotic SECIS, and in particular the conservation of the kink-turn region, we devised a thorough phylogeny-based search for SBP2 homologues in Lokiarchaeota (see Methods). We used a L7AE protein profile to identify all proteins containing this domain both in the Laz proteome, and in all eukaryotic and archaeal proteomes available at NCBI. After reducing the set to workable size by automatic selection of representative sequences, we ran our phylogenetic reconstruction pipeline. In the resulting phylogenetic tree (Supplementary Material S4.E), all known SBP2 and SBP2-like sequences clustered together, enabling us to classify as SBP2 all the genes in this cluster. This procedure identified known SBP2 genes throughout all eukaryotic lineages, including the very diverse protozoans and also nematodes, which possess an elusive, very short isoform (Otero et al., 2014). Despite our efforts, however, we could not detect any SBP2 candidate in any Laz (or Loki) sequences, or in any other archaea.

Discussion

Selenoproteins are synthesized through a conserved UGA-recoding mechanism, considered an expansion of the genetic code (Böck et al., 1991). Living organisms use this intriguing strategy to insert the amino acid Sec in catalytic sites of certain oxidoreductases. Although Sec was likely present in the last universal common ancestor, and has since been preserved in organisms in the three domains of life, including humans, it is absent in a considerable fraction of extant organisms. Although approximately half of sequenced eukaryotes contain
selenoproteins, these proteins are found only in ~20% of sequenced bacterial genomes (Lin et al., 2015; Mariotti et al., 2015), and Sec utilization is even more scarce within archaea: selenoproteins were so far detected only in two archaeal orders, *Methanopyrus* and *Methanococcales*, comprising ~12% of currently available sequenced archaeal genomes. In this work, we describe the discovery of the Sec trait in a novel archaeal phylum, *Lokiarchaeota*, the closest archaeal relative to eukaryotes known to date (Spang et al., 2015). The identity and functions of the selenoprotein genes we identified is backed by the monophyly with their experimentally characterized orthologues. Our results show that *Lokiarchaeota* possess the full set of genes to support Sec biosynthesis and incorporation, as well as selenoprotein genes belonging to 5 different families (*SPS*, *VhuU*, *VhuD*, *HdrA*, and *PrxL*, Figure 3). All selenoprotein families identified in *Loki*, except one, were previously identified in other archaeal lineages (Stock and Rother, 2009). The only exception, *PrxL*, belongs to the thioredoxin-like superfamily, like many other selenoproteins, and it was reported in other computational studies both in bacteria (Zhang and Gladyshev, 2008) and eukaryotes (Jiang et al., 2012; Mariotti et al., 2013).

We noticed that some of the *Loki* selenoproteins carry a rather unusual number of Sec residues. Whereas the known Prx and Prx-like selenoproteins have a single Sec site, we observed two in *Loki PrxL.1*, as well as in some of its homologues in the *Laz* assembly (Supplementary Table S1). The Sec residues are found in a redox box, suggesting that the *PrxL.1* protein forms a diselenide bond, a feature that has only one additional known case (Shchedrina et al., 2007). *HdrA* has also never been observed with more than one Sec residue, but in *Loki* we found such genes containing up to four Sec sites, located in five possible positions (Figure 3). *HdrA* is FAD-dependent protein with four [4Fe-4S] clusters coordinated by strictly conserved Cys residues. All Sec sites align to Cys positions that are conserved across the entire *HdrA* family, but that are not the ones involved in cluster coordination, with the only exception of the most C-terminal Sec in *HdrA.2* (Figure 3). This Sec residue is aligned with one of the four Cys in the last [4Fe-4S]
binding motif; however, Sec is here followed by an additional Cys right next to Sec, so that the motif contains four Cys and one Sec, instead of the standard four Cys residues. \( HdrA \) functions as an electron-transfer relay in the Hdr complex, providing reducing power to the catalytic subunit \( HdrB \) (Hedderich et al., 2005). The presence of several Sec residues in \( Loki HdrA \) would indicate that \( HdrA \) possesses multiple catalytic Cys residues. This suggests an unusual redox chemistry that has not been explored for this protein, traditionally considered as an intermediate in electron transfer.

The most intriguing discovery in our analysis was that the \( Lokiarchaeota \) selenoprotein genes possess a eukaryotic-like SECIS element. In fact, the lokiSECIS exhibits a conserved core which is essentially identical to the eukaryotic SECIS, and also shows the same conserved stretch of adenosines in the apical loop (Figure 4). Since the core of the eukaryotic SECIS folds in a kink-turn motif (Latrèche et al., 2009), we expect the same structure from the lokiSECIS. This implies that this motif, and the general fold of the eukaryotic SECIS, had been already established in \( Lokiarchaeota \) and was later maintained in eukaryotes. Furthermore, the lokiSECIS also distantly resembles one particular SECIS element of the selenoprotein gene \( VhuD \) in several archaea. Remarkably, all \( Methanococcales \) and \( Methanopyrus VhuD \) SECIS elements conserve the core motif AUGA preceding the main stem, which is not found in the rest of SECIS elements in the same genomes. This suggests that the archaeal \( VhuD \) SECIS is the prototypical SECIS element, and that every lokiSECIS, and by extension every eukaryotic SECIS element, descended from it.

Further support to this idea comes from the analysis of \( VhuD \) and its genomic neighbor \( HdrA \). In an unprecedented configuration, in \( Lokiarchaeota \) these two genes share a single lokiSECIS located downstream of \( VhuD \), and thus are most likely translated from the same polycistronic mRNA. In addition, these genes always co-occur with the same genomic organization (Figure 3)
in *Lokiarchaeota* (Supplementary Material S1). In every single case, *HdrA* is upstream of *VhuD*, with the same orientation and extremely short intergenic distance. Remarkably, it seems that the single lokiSECIS in *HdrA-VhuD* can support Sec incorporation for up to six UGA codons, which span a distance of ~2 Kb. This would be unparalleled in the tree of life. Outside *Lokiarchaeota*, very few selenoprotein genes with more than one Sec residue are known: *SelP* (vertebrates) (Hill et al., 1993; Lobanov et al., 2008), *SelL* (some metazoans) (Shchedrina et al., 2007), *MsrB* of *Metridium senile* (cnidarian) (Lee et al., 2011), and the *VhuD* homologue in other Sec utilizing archaea. *SelP* and *M. senile MsrB* have several Sec residues, and possess two SECIS elements each. *SelL* possess a single SECIS, which acts on two UGA codons in very close proximity (2 codons apart). The *Methanococcales* and *Methanopyrus VhuD* has been the record holder for most distant Sec residues inserted by a single (archaeal) SECIS (50 codons apart). In *Lokiarchaeota*, the *VhuD* gene extended this pattern to its neighbouring gene, *HdrA*. Plausibly, the peculiar features of the archaeal *VhuD* SECIS were advantageous in *Lokiarchaeota*, so that it took over the role of the neighboring *HdrA* SECIS, which degenerated. Ultimately, the *VhuD* SECIS ‘dictated’ its characteristics to the rest of SECIS elements, becoming effectively the new prototype structure for Sec insertion sequences in *Lokiarchaeota*, and then eukaryotes. The AUGA - GA motif in particular, which is conserved in the SECIS of archaeal *VhuD*, but not in other genes, spread to all selenoprotein genes and became what is known today as the SECIS core.

By examining the identity of the stop codons at the end of the *Lokiarchaeota* selenoprotein coding sequences, we noticed that many of them are UGA codons (Supplementary Table S1). Since lokiSECISes are located in close proximity to stop codons, structural constraints would preclude SECIS function for these positions. Thus, it appears that the lokiSECIS is able to support Sec incorporation in distant UGA codons, but has a minimal distance requirement, analogously to the eukaryotic SECIS (Martin et al., 1996; Labunskyy et al., 2014). This model
finds support in the observation that, while the various VhuD genes bear diverse stop codons including UGA, HdrA always carries UAA or UAG. We reasoned that, if this stop codon was a UGA, it would be also translated as Sec, and this mutation is selected against. It appears that the lokiSECIS can overlap the coding sequence, at least partially (Figure 3). In the VhuU.1 gene, the overlap is particularly extensive, so that its AUGA motif actually works also as the stop codon for this gene (UGA).

Despite our efforts, we could not detect SBP2 in Loki. Its apparent absence suggests that the transformation of the SECIS predates the origin of this protein. A recent study (Seeher and Schweizer, 2014) showed that, surprisingly, the deletion of SBP2 in mice neurons did not fully abolish selenoprotein expression. It is tempting to make a connection between this evidence of SBP2-independent Sec insertion in mammals and the situation in Loki, where eukaryotic-like SECIS elements are present but no SBP2 could be detected. However, the SBP2-independent selenoprotein expression in mouse may also be explained by the presence of SBP2L, a L7AE-domain containing paralog of SBP2 that emerged at the root of vertebrates, that was previously shown to weakly bind SECIS elements in vitro (Donovan and Copeland, 2009). Unfortunately, the question of archaeal SECIS recognition remains, to date, unanswered. Like for the rest of Sec utilizing archaea, we suppose that the SBP2 function in Lokiarchaeota is either carried out by an unknown dedicated protein, or performed by a constitutive ribosomal component (e.g. L30) as an accessory function. Given the different SECIS structures in Lokiarchaeota and other Sec encoding archaea, it is plausible that the protein responsible for their binding is distinct in the two cases.

In conclusion, our work sheds light on the transition of the Sec recoding pathway from archaea to eukaryotes, testifying the value of the Lokiarchaeota genome as a “living fossil” between prokaryotes and eukaryotes. The selenoprotein genes in Lokiarchaeota are mostly typical of the
archaeal world, but they possess conserved RNA structures with unmistakable similarity to eukaryotic SECIS elements.

Materials and methods

5.1 Identification of selenoprotein and Sec machinery genes

We searched the Loki sequences for selenoprotein genes using a combination of computational methods. First, we applied Selenoprofiles, a homology-based pipeline able to correctly predict known selenoprotein families in genomes even in complete automation (Mariotti and Guigó, 2010). Second, we ran blast searches (Altschul et al., 1997) (tblastn program) using a comprehensive set of selenoproteins annotated in other species, and further manually inspected the results. Third, we used a modified version of Seblastian (Mariotti et al., 2013). This program identifies potential SECIS elements as a first step, and then searches for known or novel selenoproteins in the sequence upstream of each SECIS, using blastx against a database of known selenoproteins or potential Cys homologues. For Loki, due to the modest total sequence size, we could apply a modified Seblastian that bypassed SECIS finding and searched the full metagenome instead. All candidates from the three approaches were merged in a single set, and were subjected to extensive manual analysis and gene structure refinement. This resulted in a set of 13 UGA-containing Loki selenoprotein genes, belonging to the families HdrA, VhuD, VhuU and PrxL (Supplementary Table S1). Analyzing their genomic neighborhoods (see “synteny analysis” below), we reduced this to a non-redundant set of 10 UGA-containing selenoprotein genes. This set allowed the first discovery of the lokiSECIS motif, which was then progressively enriched with homologous sequences in Laz to form a lokiSECIS model (see “search for Lokiarchaeota SECIS” below). Performing searches with this model, we obtained a (potentially redundant) set of 33 selenoproteins in Laz (a superset of the Loki selenoprotein set), which all belong to aforementioned protein families. In addition to these genes, we identified ten
Loki-like SPS selenoproteins in Laz. Furthermore, the Laz sequence set contained many other selenoprotein sequences attributed to a variety of bacterial species, which were filtered out. All selenoprotein sets were subjected to phylogenetic analysis (Supplementary Material S1). Sec machinery protein coding genes (SecS, EFsec, pstk) were identified using Selenoprofiles and manual tblastn searches, followed by phylogenetic analysis to ensure the correct assignment of gene family (Supplementary Material S3). The search for SBP2 was carried out with a particularly exhaustive procedure (see “search for SBP2” below). tRNAsec was identified using a newly developed method based on covariance models, which was built specifically to find this gene in nucleotide sequences [Santesmasses, in preparation].

5.2 SECIS models for archaea and eukaryotes

For archaea, a first structural alignment of a few SECIS elements was manually assembled based on known sequences of archaeal selenoprotein genes (Kryukov and Gladyshev, 2004). This first “seed” model was subsequently enriched with additional archaeal SECISes, in the following way. From all archaeal genomes available at NCBI, we selected those that, based on the presence of Sec machinery (analogously to (Mariotti et al., 2015)), were predicted to code for Sec. All such genomes belonged to the Methanococcales and Methanopyrus orders. We used Selenoprofiles to identify their selenoprotein genes, and finally scanned their downstream sequences with the seed model, using the program infernal (Nawrocki and Eddy, 2013). The final set of potential archaeal SECISes was inspected and filtered to obtain a bona fide set (71 sequences). The criteria chosen for filtering were the distance with the coding sequence and the fit with the seed consensus structure. This archaeal SECIS model was used for two purposes. First, it was used to search the Loki selenoproteins for archaeal SECISes, which returned no hits. Second, it was used to obtain a graphical representation of the archaeal SECIS consensus (Figure 4). To this aim, we removed all columns with >85% gaps, and the resulting alignment was run with weblogo (frequency plot) (Crooks et al., 2004) and RNAalifold (Bernhart et al.,
The images obtained in this way were assembled and colored to produce the scheme shown in Figure 4.

For eukaryotic SECISes, searches were performed using the program SECISearch3 (Mariotti et al., 2013), set at best sensitivity. To obtain a graphical representation of the eukaryotic SECIS consensus, we used the structural alignment underlying SECISearch3. This large set (1121 sequences) was reduced by selecting the ~250 most representative sequences using trimal (Capella-Gutiérrez et al., 2009), and further trimmed by removing all columns with >25% gaps. The resulting alignment was processed with weblogo and with RNAalifold to produce the two graphical components (frequency logos and structure) then assembled in Figure 4. For aesthetic reasons, the non-canonical pairing of the two GA dinucleotides at the kink-turn core was imposed as constraint when running RNAalifold.

5.3 Search for the Lokiarchaeota SECIS and RNA structure prediction

Initially, we searched the Loki selenoprotein gene sequences using SECIS covariance models (Nawrocki and Eddy, 2013) from archaea, eukaryotes (see above), and bacteria [Santesmasses, unpublished], but this gave no significant matches. We then set out to search for motifs de novo. Our sequence dataset for motif search consisted in the non-redundant Loki selenoprotein set with the addition of the Loki SPS fragment (Supplementary Table S1). We considered three sequence classes: the coding sequences, the regions just upstream, and those just downstream. We exploited the NCBI annotation of Loki sequences to delimit the upstream and downstream regions, by cutting just before the start or end of the next annotated gene. Regions that were shorter than 20 nucleotides were excluded from all subsequent analyses, removing in the process the small intergenic regions between the HdrA-VhuD pairs. All remaining upstream and downstream regions were further extended by 30 nucleotides towards their gene, enabling the motifs to have a partial overlap with coding sequences. We
then ran the motif search program Glam2, able to predicted motifs potentially containing gaps (Frith et al., 2008), on the three sequence sets separately (coding sequences, upstream, downstream). To assess significance of the predicted motifs, we searched for them again in the full genome and examined the locations and scores of their occurrences along the *Loki* selenoprotein genes, as well as their predicted structure and free energy. The best scoring motif in the downstream sequences appeared to be a suitable candidate: in contrast to all other motifs, it folded consistently in very similar shapes and all localized in homologous positions, in close proximity to the putative translation termination site. The occurrences of this motif were used to train a first “lokiSECIS” infernal model (Nawrocki and Eddy, 2013), using RNAalifold (Bernhart et al., 2008) to predict a consensus RNA structure and emacs ralee (Griffiths-Jones, 2005) to inspect it. A pattern expression for the program scan_for_matches (http://blog.theseed.org/servers/2010/07(scan-for-matches.html) was also manually designed to fit the occurrences of this motif in *Loki* selenoproteins. We then built a “lokiSECISearch” program. Inspired by the original SECISearch algorithm (Kryukov et al., 1999), this program uses both the motif pattern and the infernal model to search nucleotide sequences. It then predicts the structure of the hits using RNAfold (Lorenz et al., 2011), and filters out those with free energy greater than -5.0 kcal/mol. Finally, the program reports any UGA-containing ORFs located upstream of SECIS candidates. We ran lokiSECISearch on the *Loki* and *Laz* metagenomes. While no further selenoprotein candidate was predicted in *Loki*, after filtering and manual curation we obtained a set of 33 selenoprotein genes in *Laz*, all belonging to the families *HdrA, VhuD, VhuU, PrxL*. We used the downstream motif occurrences in these genes to enrich our lokiSECIS model, which finally consisted of 25 sequences. To produce a graphical representation like for the archaeal and eukaryotic models, the lokiSECIS alignment was trimmed removing columns with >85% gaps, processed with weblogo and RNAalifold (imposing the pairing of the GA dinucleotides at the core), and finally colored and assembled (Figure 4).
With our enriched lokiSECIS model, we could now detect a match downstream of each single *Loki* selenoprotein gene (for *HdrA-VhuD* pairs, the lokiSECIS was located only downstream of the second gene). Notable exceptions were the *Loki* SPS fragment, as well as the *Loki*-like SPS in *Laz*, which did not exhibit this motif. At this stage, however, due to searches in *Laz*, we had at hand an adequate number of *Loki*-like selenoprotein sequences to attempt a different *de novo* approach to find conserved structures. For each selenoprotein family separately (*HdrA*, *VhuD*, *VhuU*, *PrxL*, and *SPS*), we extracted the genes’ coding sequences extended by 200 nucleotides at each side and aligned them using clustal omega (Sievers et al., 2011). We then used the program RNAz (Gruber et al., 2010) to detect any family-specific conserved structure supported by compensatory mutations. RNAz reported significant hits corresponding to every lokiSECIS element predicted so far (downstream of *PrxL*, *VhuD* and *VhuU* genes). Furthermore, it predicted a stable structure downstream of the *Loki*-like *SPS* genes. This structure, although exhibiting obvious similarities to the lokiSECIS of the other *Loki* selenoprotein genes, has also several important differences (see Results and Figure 5), which make it evade the detection power of both the enriched lokiSECIS infernal model and patscan pattern. RNAz did not report any significant hits other than those just described. We built an additional infernal model for the SPS lokiSECIS structure, and incorporated it in the program lokiSECISearch.

5.4 Synteny analysis

We set out to find homologous relationships in the genomic context of our genes of interest (the selenoproteins and Sec machinery in *Loki* and *Laz*). In order to do that, we downloaded the NCBI protein annotations of *Loki* and *Laz*, and we integrated them with our curated selenoprotein gene sets (since selenoproteins were not correctly annotated in NCBI). The resulting protein set was run with blastp against itself, and partitioned in protein families by single link clustering based on the blastp matches with an e-value lower than $10^{-12}$. We then used the syntheny_view program (available at https://github.com/marco-mariotti/tree_classes), a
ETE2-based (Huerta-Cepas et al., 2010) script that produces a graphical representation of gene surrounds, highlighting with the same color the genes that belong to the same family. With this visualization, it became evident that several genes were present in multiple copies with an homologous gene context (Supplementary Material S1). We considered these to be different “versions” of the same gene (i.e., orthologues in closely related species, or just variants in a population), and based on this we reduced our initial Loki gene set to a non-redundant set (Supplementary Table S1).

5.5 Search for SBP2

Our initial searches for SBP2 in Loki were performed using Selenoprofiles (Mariotti and Guigó, 2010) and tblastn of known eukaryotic homologues, but gave no suitable candidates. We thus performed a more thorough search, with the following procedure. We downloaded all archaeal and eukaryotic NCBI genome assemblies and extracted their protein coding annotation using the Entrez module of Biopython (Cock et al., 2009) and GBParsy (Lee et al., 2008). We then scanned all the proteomes obtained in this way using hmmer version 3.1b1 (Eddy, 2009), with the pfam profile (Finn et al., 2016) of the L7AE domain (PF01248.22), which is conserved in all SBP2 proteins but also shared with other ribonucleoproteins. We performed the same hmmer search on all proteins annotated in the Laz sequences. We extracted the protein sequences of all domains matching this profile with an e-value lower than 0.01 (4,527 sequences), and we aligned them using the program mafft v7.215 (Katoh and Standley, 2013). We then used trimal (Capella-Gutiérrez et al., 2009) to select the best representative sequences with a maximum sequence identity of 92%, obtaining a reduced set of 1,962 proteins with a L7AE domain. Finally, we ran our phylogenetic reconstruction pipeline (see below) on this alignment, and we inspected the resulting tree (Supplementary Material S4.E). The sequences consistently clustered by their annotated family, with known SBP2 proteins (including here the vertebrate
SBP2-like subfamily (Donovan and Copeland, 2009)) falling in a single cluster. We concluded that all domains in this cluster, and only these, belonged to bona fide SBP2.

5.6 Phylogenetic analysis

Phylogenetic trees were computed by maximum likelihood with the evolutionary model resulting from an automated selection procedure, as explained in (Mariotti et al., 2012) after (Huerta-Cepas et al., 2011). The input were protein sequence alignments generated by mafft v7.215 (Katoh and Standley, 2013) or clustal omega v1.2.1 (Sievers et al., 2011). For each protein family of interest (selenoproteins and Sec machinery), we included in the alignment their most similar proteins in NCBI NR, identified using blastp. The e-value threshold was manually adjusted for each family, making sure that searches were permissive enough to include also similar, but non-orthologous proteins (e.g. EF-TU sequences were included in the EFsec alignment). Trimal (Capella-Gutiérrez et al., 2009) was run to remove very similar sequences (>90% identity) and also to trim non-informative columns using a method optimized for maximum likelihood (-automated1 option, see trimal manual). For SPS alone, instead of collecting homologous sequences with blastp, we used the manually curated set in (Mariotti et al., 2015).

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References


Figures legends

**Figure 1. Sec machinery in the three domains of life.** The figure shows two modules of the Sec pathway: Sec biosynthesis (A), and Sec insertion during selenoprotein translation (B). The differences between bacteria, archaea and eukaryotes (Yoshizawa and Böck, 2009; Stock and Rother, 2009; Labunskyy et al., 2014) are marked with different colors. SerRS: serine-tRNA synthetase; pstk: phosphoseryl-tRNA kinase; SecS: archaeal/eukaryotic Sec synthase; SelA: bacterial Sec synthase; SPS: selenophosphate synthetase; SelB: bacterial/archaeal Sec elongation factor; eEFsec: eukaryotic Sec elongation factor; SBP2: SECIS binding protein 2; bSECIS: bacterial SECIS; aSECIS: archaeal SECIS; (SECIS: eukaryotic SECIS); SRE: Sec redefinition element (Howard et al., 2007).

**Figure 2. tRNAsec in Lokiarchaeota.** The color scheme highlights the tRNA stems: red for the acceptor arm, yellow for the D arm, green for the anticodon arm, blue for the variable arm, and
purple for the T arm. This tRNA has all the characteristics expected for an archaeal tRNAsec: UCA anticodon, G discriminator base, 9/4 fold of the acceptor/T stems, a D arm of 7 bp, a D loop of 4 bp, and a long variable arm (Chiba et al., 2010). Two copies of tRNAsec with very similar sequence were identified in Loki (Supplementary Table S1), but since their gene context is identical (Supplementary Material S1.A), we conclude that they correspond to the same, sole copy of this gene in Lokiarchaeota.

**Figure 3. Selenoprotein genes in Lokiarchaeota.** The VhuU.1 gene (first on top) serves as legend. Only a fragment of the SPS gene was identified in Loki, but the missing sequence could be deduced from similar gene occurrences in Laz. The lokiSECIS in SPS is colored differently to indicate its peculiar characteristics (Figure 4 and 5). Note that the coding sequence of genes HdrA.3 and VhuD.3 overlap in different frames by 28 nucleotides. The genes HdrA.4/VhuD.4, PrxL.1 and VhuU.1 are present in one extra copy in the Loki contigs (Supplementary Table S1), but these are not shown here since their gene context is identical (Supplementary Material S1.A).

**Figure 4. SECIS elements in Lokiarchaeota compared with eukaryotes and other archaea.** SECIS elements in known Sec utilizing archaea (A), archaeal VhuD genes (B), Lokiarchaeota (see also Figure 5) (C), and eukaryotes (D). These images were derived from curated SECIS alignments as explained in Methods. The structures are predicted by RNAalifold (Bernhart et al., 2008) and incorporate the frequency logos (Crooks et al., 2004) of the nucleotides found at each position.

**Figure 5. A distinct SECIS element in Loki SPS.** The figure shows the SECIS predicted downstream of the Loki SPS fragment. This structure, predicted by RNAfold (Lorenz et al., 2011), is analogous to that predicted for all other Loki-like SPS in Laz, and also to their
consensus structure predicted by RNAalifold (Supplementary Material S4.C). The figure highlights the features shared with the rest of *Loki* SECIS and the eukaryotic SECIS (Figure 4). Those nucleotide positions conserved in all *Lokiarchaeota* SECISes, but not in eukaryotes, are circled in blue.
**Sec biosynthesis pathway**

- **bacteria**: brown
- **archaea**: blue
- **eukarya**: orange

**A**

- Ser-tRNA<sub>sec</sub> → phoshoSer-tRNA<sub>sec</sub> → Sec-tRNA<sub>sec</sub>
- SerRS → Ser-tRNA<sub>sec</sub>
- SPS → SelA
- HSe<sup>-</sup> → Se<sup>P</sup>

**B**

**Bacteria**

- START → UGA → STOP
- tRNA<sub>sec</sub>
- Sec-tRNA<sub>sec</sub>
- SelB
- bSECIS

**Archaea**

- START → UGA → STOP
- tRNA<sub>sec</sub>
- Sec-tRNA<sub>sec</sub>
- SelB
- aSECIS
- 3' UTR

**Eukaryotes**

- START → UGA → STOP
- tRNA<sub>sec</sub>
- Sec-tRNA<sub>sec</sub>
- eEF<sub>sec</sub>
- SBP2
- SECIS
- SRE
- 3' UTR